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APPLICATION NOTE 70923

Sensitive and rapid determination of polycyclic aromatic hydrocarbons in tap water

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Keywords

Hypersil Green PAH Column, Acclaim PA2 Column, HPLC, On-Line SPE, PAHs, UV Detection, EPA Method 550, EPA Method 550.1, Water Analysis

Goal

To develop an efficient high-performance liquid chromatography (HPLC) method for sensitive and rapid determination of 20 polycyclic aromatic hydrocarbons (PAHs) in environmental waters using on-line solid-phase extraction (SPE) for sample preparation instead of the liquid-liquid extraction and off-line SPE specified in the U.S. Environmental Protection Agency (EPA) Methods 550 and 550.1, respectively

Introduction

Polycyclic aromatic hydrocarbons (PAHs) are a group of chemicals formed from the incomplete combustion of organic matter. Due to their potential carcinogenic and mutagenic properties, most countries have regulations limiting the concentrations of a variety of PAHs in drinking water, food additives, cosmetics, workplaces, and factory emissions.

Thermo Fisher Scientific Application Note (AN) 196 provides on-line SPE HPLC methods to quantify low concentrations of PAHs in oil.[1] The costs for the SPE cartridge, labor, time, and reagents are significantly reduced using these methods, and the results are more consistent. This is because on-line SPE eliminates manual processes, such as rotary evaporation and nitrogen-assisted evaporation in the routine liquid-liquid extraction and off-line SPE steps described in EPA Methods 550, 550.1, and 610.[2-4] However, because the run time of the reported on-line SPE HPLC method exceeded 60 min in this study, the analytical column used in the AN was replaced to shorten analysis time.

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Equipment, Software, and Consumables

- Thermo Scientific™ UltiMate™ 3000 Dual Gradient Rapid Separation (RS) LC system, including:
  - Thermo Scientific™ UltiMate™ 3000 DGP-3600RS Dual Gradient Rapid Separation Pump (P/N 5040.0066)
  - Thermo Scientific™ UltiMate™ 3000 SRD-3600 Integrated Solvent and Degasser Rack (P/N 5035.9230)
  - Thermo Scientific™ UltiMate™ 3000 WPS-3000TRS Rapid Separation Wellplate Sampler, Thermostatted (P/N 5840.0020) with a 1000 µL sample loop (P/N 6820.2429) and a 1000 µL syringe (P/N 6822.0005)
  - Thermo Scientific™ UltiMate™ 3000 TCC-3000SD Standard (P/N 5730.0010) or TCC-3000RS Rapid Separation (P/N 5730.0000) Thermostatted Column Compartment, equipped with one 2p–6p valve
  - Thermo Scientific™ UltiMate™ 3000 DAD-3000RS Rapid Separation Diode Array Detector (P/N 5082.0020)
  - Thermo Scientific™ UltiMate™ 3000 FLD-3400RS Rapid Separation Fluorescence Detector (P/N 5078.0025)
- Thermo Scientific™ Chromeleon™ Chromatography Data System (CDS) software, version 7.1 or above
- Thermo Scientific™ Target2™ Nylon Syringe Filters, 0.45 µm, 30 mm (P/N F2500-1)

Reagents and Standards

- Deionized (DI) water, 18.2 MΩ-cm resistivity
- Methanol (CH_3OH), 99.8%, HPLC Grade (Fisher Scientific P/N AC610090040)
- Acetonitrile (CH_3CN), HPLC Grade (Fisher Scientific P/N AC610010040)
- EPA 610 PAH Solution (1000 mg/L for each, in methylene chloride) containing 18 PAH standards: benzo[k]fluoranthene, acenaphthene, acenaphthylene, anthracene, fluorene, naphthalene, phenanthrene, benzo[a]anthracene, benzo[a]pyrene, chrysene, fluoranthene, indeno[1,2,3-cd]pyrene, pyrene, benzo[b]fluoranthene, benzo[ghi]perylene, dibenz[a,h]anthracene, benzo[e]pyrene, and benzo(j)fluoranthene (o2si Smart Solutions P/N 110064-01-5PAK)
- 1-Methylnaphthalene (AccuStandard P/N H-001N)
- 2-Methylnaphthalene (AccuStandard P/N H-002N)

Conditions (Applicable to Figures 1, 2, and 6)	
On-Line SPE	
Column:	Thermo Scientific™ Acclaim™ PolarAdvantage II (PA2), 3 µm Analytical, 4.6 × 50 mm (P/N 063189)
Mobile Phase:	A. Water B. Acetonitrile
Gradient:	Table 1
Flow Rate:	0.4 and 0.6 mL/min (Table 1)
Inj. Volume	1 mL on the On-Line SPE column
Separation	
Column:	Thermo Scientific™ Hypersil™ Green PAH Column, 3 µm, 3.0 × 150 mm (P/N 31105-153030)
Mobile Phase:	A. Water B. Acetonitrile
Gradient:	Table 1
Flow Rate:	0.8 mL/min
Temperature:	30 °C
Detection:	UV, 254 nm; Fluorescence at different excitation and emission (Ex/Em) wavelengths for each PAH (Table 2)

Note: The PAHs have good fluorescent responses, except for acenaphthylene (Figure 1, Peak 2). Because their maximum fluorescent responses occur at different Ex/Em wavelengths, it is necessary to change the Ex/Em wavelengths to acquire the best detection sensitivity. These changes are dictated by the individual PAH retention times. Table 2 shows the program for wavelength changes. Although naphthalene (Figure 1, Peak 1) has fluorescent response, EPA method 550.1 requires it to be determined using UV detection, together with acenaphthylene (Figure 1, Peak 2). Figures 1 and 2 show the chromatograms of all 20 PAHs with UV and fluorescence detection, respectively, under the conditions specified above.

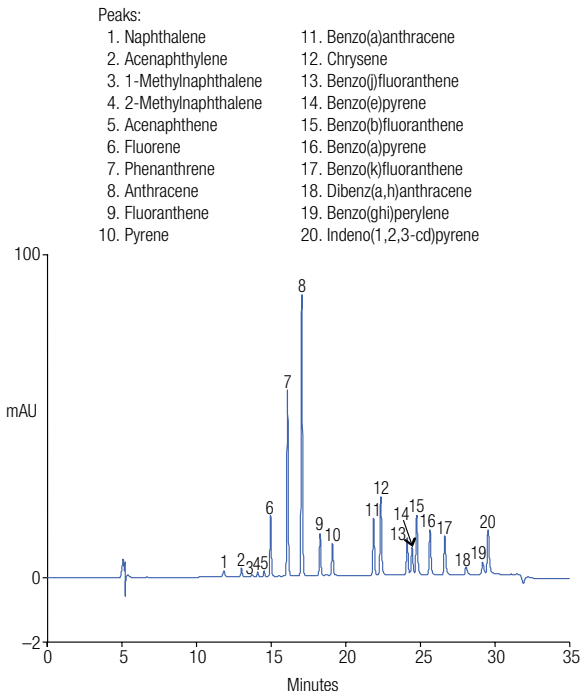

Figure 1. All 20 PAHs (50 µg/L for each PAH) detected at UV 254 nm.

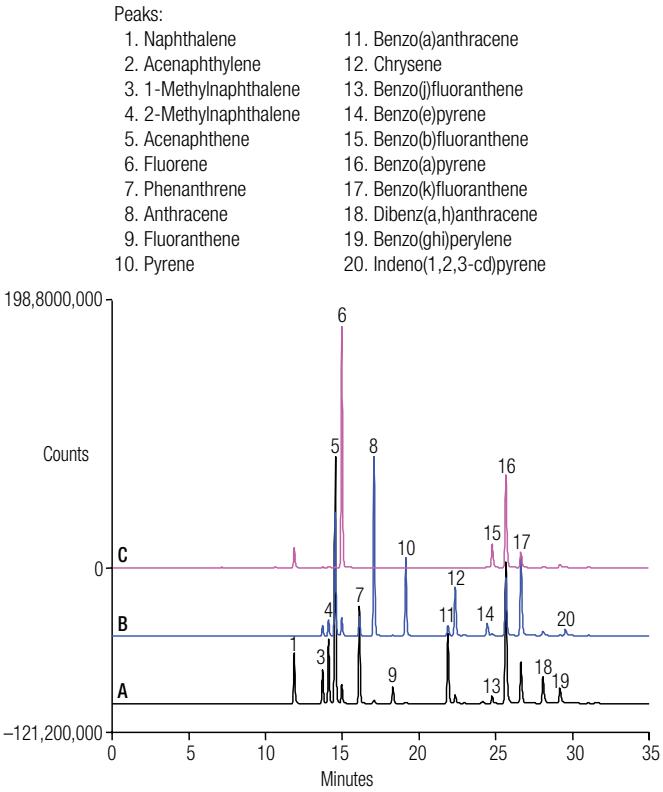

Figure 2. All 20 PAHs (50 µg/L for each PAH) detected by fluorescence detection using programmed wavelength switching in three parallel channels: (A) Emission_1, (B) Emission_2, (C) Emission_3. Note: Acenaphthylene (Peak 2) is not shown here because there was no fluorescent response for acenaphthylene.

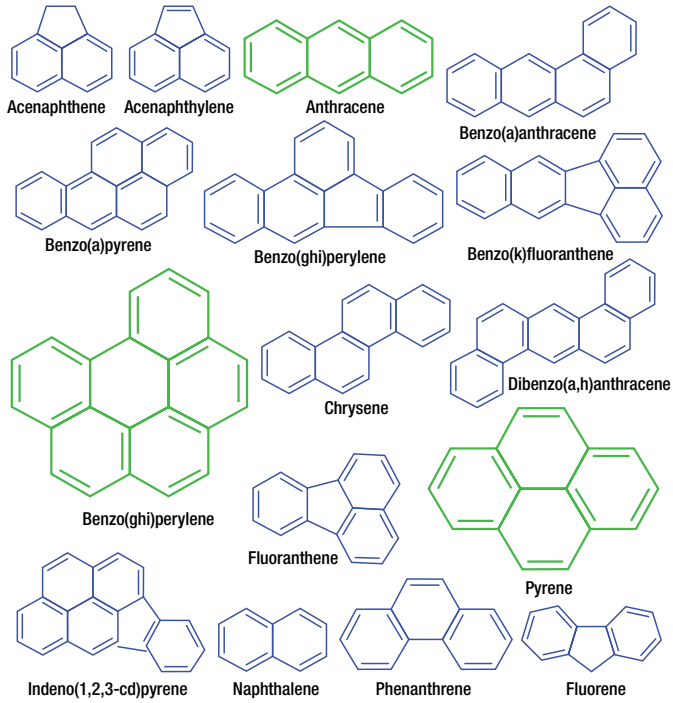

Figure 3. Structures of the 16 PAHs specified in EPA Methods 550, 550.1, and 610.

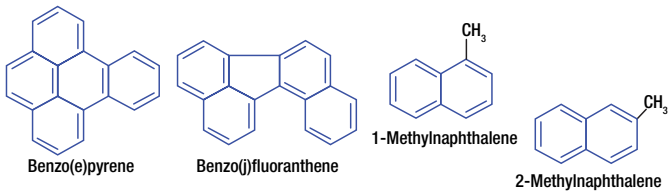

Figure 4. Structures of four additional PAHs not specified in EPA Methods 550, 550.1, and 610.

Preparation of Standard Solutions

In addition to the 16 PAHs specified in EPA Methods 550, 550.1, and 610 (Figure 3), the target analytes in the experiments described here include four other PAHs: benzo[e]pyrene, benzo(j)fluoranthene, 1-methylnaphthalene, and 2-methylnaphthalene (Figure 4). The EPA 610 PAH Solution product contains benzo[e]-pyrene and benzo(j)fluoranthene, in addition to the 16 specified PAHs.

Stock Solutions of 1-Methylnaphthalene and 2-Methylnaphthalene

In a 100 mL volumetric flask, dissolve 100 mg of 1-methyl-naphthalene in 2 mL of acetonitrile and dilute to the mark with methanol. The final concentration of 1-methylnaphthalene will be 1000 mg/L. Prepare a 1000 mg/L stock solution of 2-methylnaphthalene in the same manner.

Stock Standard Mixes 1 and 2

Add 100 µL of EPA 610 PAH Solution (containing 18 PAH standards, 1000 mg/L each), 100 µL of 1-Methylnaphthalene Stock Solution, and 100 µL of 2-Methylnaphthalene Stock Solution to a 10 mL volumetric flask, then dilute to the mark with methanol. The final concentration of each PAH will be 10 mg/L. This is Stock Standard Solution Mix 1.

Add 1 mL of Stock Standard Mix 1 to a 10 mL volumetric flask and dilute to the mark with methanol. This is Stock Standard Solution Mix 2. The final concentration of each PAH in Stock Standard Mix 2 will be 1.0 mg/L.

Use these mixed standard stock solutions to prepare working mixed standard solutions for calibration.

Working Mixed Standard Solutions for Calibration

For calibration, prepare nine working standard solutions with different concentrations by diluting the proper amount of either Stock Standard Mix 1 or 2 with DI water. The volumes of each solution needed to make the calibration standards are shown in Table 3. The

Table 1. Gradient programs for loading and analytical pumps.

Loading Pump				Analytical Pump			
Time (min)	Flow Rate (mL/min)	% A (H_2O)	% B (CH_3CN)	Time (min)	Flow Rate (mL/min)	% A (H_2O)	% B (CH_3CN)
0	0.6	95	5	0	0.8	60	40
4.0	0.6	95	5	5	0.8	60	40
4.5	0.4	0	100	30	0.8	0	100
25	0.4	0	100	30.5	0.8	60	40
25.5	0.6	95	5	35	0.8	60	40
35	0.6	95	5	—	—	—	—

Table 2. Ex/Em maximums for each PAH and programmed wavelength switching times.

Time (min)	Fluorescence Detection Channel	Ex/Em Wavelengths (nm)	PAH	Peak No.
0.0	Emission_1	219/330	Naphthalene	1
13.45	Emission_1	225/333	1-Methylnaphthalene	3
			2-Methylnaphthalene	4
	Emission_2	235/332	Acenaphthene	5
	Emission_3	263/310	Fluorene	6
15.50	Emission_1	247/364	Phenanthrene	7
	Emission_2	247/401	Anthracene	8
17.80	Emission_1	281/453	Fluoranthene	9
	Emission_2	236/389	Pyrene	10
20.50	Emission_1	281/391	Benzo(a)anthracene	11
	Emission_2	264/381	Chrysene	12
23.50	Emission_1	240/510	Benzo(j)fluoranthene	13
	Emission_2	283/394	Benzo(e)pyrene	14
	Emission_3	249/443	Benzo(b)fluoranthene	15
25.40	Emission_1	243/412	Benzo(k)fluoranthene	16
	Emission_2	260/408	Benzo(a)pyrene	17
27.50	Emission_1	290/398	Dibenz(a,h)anthracene	18
28.70	Emission_1	292/415	Benzo(ghi)perylene	19
	Emission_2	246/503	Indeno(1,2,3-cd)pyrene	20

Table 3. Preparation of calibration standards.

Stock Standard of PAHs Calibration Mixture	Volume of Stock Standard of PAHs Calibration Mixture (µL)	Volume of Water (mL)	Final Volume of Calibration Standard (mL)	Final Concn of Calibration Standard (µg/L)
Stock Standard Mix 1 (10 mg/L)	100	9.9	10.0	100
	50	9.95		50
Stock Standard Mix 2 (1 mg/L)	100	9.9		10
	50	9.95		5.0
Calibration Standard (100 µg/L)	100	9.9		1.0
	50	9.95		0.5
Calibration Standard (10 µg/L)	100	9.9		0.1
	50	9.95		0.05
	10	9.99		0.01

calibration standards with concentrations of 100 and 10 µg/L are also used as stock standards for the preparation of the working mixed standard solutions with lower concentrations.

Preparation of Water Samples

Tap water samples were collected from local water sources in the Pudong District, Shanghai, People's Republic of China.

Filter water samples using nylon syringe filters prior to injection.

Add 200 µL of Stock Standard Mix 2 (1.0 mg/L of each PAH) and 39.8 mL of each filtered water sample to a conical flask with plug. The concentration of each PAH in the spiked water sample will be 5 µg/L.

Add 200 µL of the calibration standard with a concentration of 10 µg/L of each PAH and 39.8 mL of each filtered water sample to a conical flask with plug. The concentration of each PAH in the spiked water sample will be 0.05 µg/L.

Results and Discussion

Evaluation of On-Line SPE

Figure 5 shows a typical flow schematic of an on-line SPE system that is directly coupled to the HPLC column using one 6-port (2p–6p) valve. The filtered sample is directly injected onto the system and delivered to the SPE column for enrichment (1_2 position) using one pump of the dual-pump module (labeled *For on-line SPE*); the analytical column is simultaneously equilibrated with the second analytical pump (labeled For Separation) of the dual-pump module. After the analytes are bound to the SPE column and impurities are washed out, the SPE column is switched into the analytical flow path to elute the bound analytes (6_1 position); then the analytes are separated on the analytical column and detected by the UV and fluorescence detectors. This method is easily accomplished using an UltiMate 3000 Dual Gradient LC system.

Selection of On-Line SPE and Analytical Columns

The Acclaim PA2 column, was chosen because of its compatibility with the water matrix, while giving high retentivity for the PAH compounds.

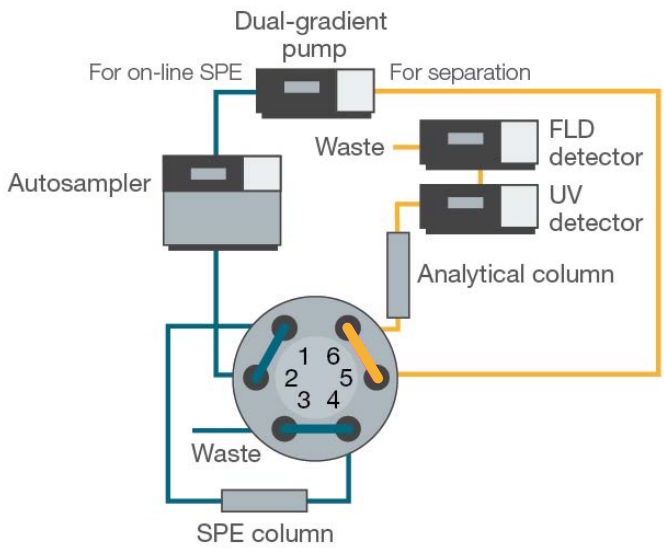

Figure 5. Flow schematic of on-line SPE.

Table 4. Calibration data and MDLs for the 20 PAHs.

Analyte	Detection	Regression Equation	r^2	Linearity Range (µg/L)	Concn of PAH Standard Mixtures in Reagent Water (µg/L)[a]	MDL (µg/L)[b]
Naphthalene	UV	A = 0.0025c + 0.0136	0.9970	1.5~100	10	0.47
Acenaphthylene		A = 0.0034c + 0.0062	0.9989	1.5~100	10	0.72
1-Methylnaphthalene	Fluorescence	A = 7492.48c	0.9910	0.10~50	0.1	0.031
2-Methylnaphthalene		A = 10516.8c	0.9982	0.10~50	0.1	0.031
Acenaphthene		A = 110163c	0.9972	0.05~50	0.1	0.028
Fluorene		A = 266343c	0.9966	0.05~50	0.1	0.016
Phenanthrene		A = 138211c	0.9966	0.05~50	0.1	0.011
Anthracene		A = 237397c	0.9973	0.05~50	0.1	0.010
Fluoranthene		A = 32560.4c	0.9971	0.05~50	0.1	0.017
Pyrene		A = 148615c	0.9965	0.05~50	0.1	0.012
Benzo(a)anthracene		A = 100842c	0.9980	0.10~50	0.1	0.020
Chrysene		A = 74195.7c	0.9986	0.10~50	0.1	0.024
Benzo(j)fluranthene		A = 2405.81c	0.9992	0.5~100	1.0	0.156
Benzo(e)pyrene		A = 13930.5c	0.9996	0.5~100	1.0	0.161
Benzo(b)fluoranthene		A = 33121.4c	0.9954	0.1~50	0.1	0.034
Benzo(k)fluoranthene		A = 213065c	0.9962	0.1~50	0.1	0.023
Benzo(a)pyrene		A = 122278c	0.9969	0.1~50	0.1	0.035
Dibenz(a,h)anthracene		A = 54922.1c	0.9997	0.1~50	1.0	0.048
Benzo(ghi)perylene		A = 22558.1c	0.9998	0.1~100	1.0	0.137
Indeno(1,2,3-cd) pyrene		A = 8615.10c	0.9979	0.1~100	1.0	0.131

[a] Used for the determination of the standard deviation value for calculating MDLs

[b] The single-sided Student's *t* test method (at the 99% confidence limit) was used for estimating MDL, where the standard deviation of the peak area of eight injections of tap water sample spiked with mixed PAHs standard is multiplied by 3.5 (at n = 8) to yield the MDL.

The Hypersil Green PAH column features a specially tailored alkyl-bonded silica with high carbon loading. This column was designed specifically for the separation of PAHs and optimized for the published EPA methods.[2–4] The column resolves benzo[e]pyrene, benzo[j]fluoranthene, and benzo[b]fluoranthene, which have similar structures (Figure 1). The structures have a different configuration, so the resolution is based on the steric selectivity of the Hypersil Green PAH column. As shown in Figures 1 and 2, using the combination of Acclaim PA2 and Hypersil Green PAH columns under the specified conditions, on-line SPE followed by HPLC can be accomplished in 35 min with baseline separation of 20 PAHs.

Determination of Ex/Em Maximums for PAHs

All 20 PAHs except acenaphthylene can be detected using a fluorescence detector, which usually offers much higher sensitivity than UV detection. As previously discussed, each PAH has its own Ex/Em maximum, and thus programmed fluorescence wavelength switching (switching to the Ex/Em maximum wavelength of each individual PAH when the PAH peak passes through the fluorescence detector) is necessary to obtain the best sensitivity for each PAH. To determine the appropriate fluorescence Ex/Em wavelength for each PAH, a PAH standard mixture was injected and the diode array detector used to obtain the maximum UV absorption (UVmax) of each PAH. This experiment showed that all UVmax of PAHs were close to 220, 240, or 280 nm. As a result, 220, 240, and 280 nm were used as the excitation wavelengths to perform emission scans for each PAH in order to determine its emission maximum. Excitation scans were performed using resultant emission maximums for all PAHs. The determined Ex/Em maximums for each PAH are shown in Table 2.

Optimization of Detection Parameters

Unfortunately, practical problems—as when two peaks elute close to one another—sometimes prevent switching to the appropriate Ex/Em wavelength for each PAH peak. When wavelength switching is programmed during the elution of a peak or even at the shoulder of a peak, the detector can be saturated and the analysis that follows can be disrupted. To resolve this problem, when two nearby peaks need to use different wavelengths, three parallel fluorescence monitoring channels are used to perform the analysis (one channel for each of the nearby peaks).

For example, as shown in Figure 2, benzo(j)fluoranthene (Peak 13), benzo(e)pyrene (Peak 14), and benzo(b) fluoranthene (Peak 15) elute in a small retention time window; therefore, three parallel fluorescence monitoring channels (Emmisions_1, _2, and _3) are used for their determination. For the same reason, 1-methylnaphthalene (Peak 3), 2-methylnaphthalene (Peak 4), acenaphthene (Peak 5), and fluorene (Peak 6) are also monitored using three parallel fluorescence monitoring channels. Table 2 lists the parallel fluorescence monitoring channels used for all 20 PAHs.

Reproducibility, Linearity, and Detection Limits

Method precision was estimated using fluorescence detection by making seven consecutive 1000 µL injections of a calibration standard with a concentration of 1 µg/L of each PAH. Method precision using UV detection was measured in the same manner, but with a calibration standard having a concentration of 10 µg/L of each PAH. Retention time reproducibilities (RSD) are all ≤0.16 and peak area reproducibilities (RSD) are all ≤1.3, thus demonstrating good short-term precision for this on-line SPE HPLC method.

Calibration linearity of 20 PAHs (using UV detection for naphthalene and acenaphthylene while using fluorescence detection for the other 18 PAHs) was investigated by making three consecutive 1000 µL injections of a mixed standard prepared at nine different concentrations (i.e., 27 total injections). The external standard method was used to establish the calibration curve and quantify the analytes in the tap water samples. Different linearity ranges were observed for the PAHs when plotting concentration versus peak area. Detailed calibration data calculated by Chromeleon CDS software are shown in Table 4. The method detection limit (MDL) of each PAH for UV or fluorescence detection

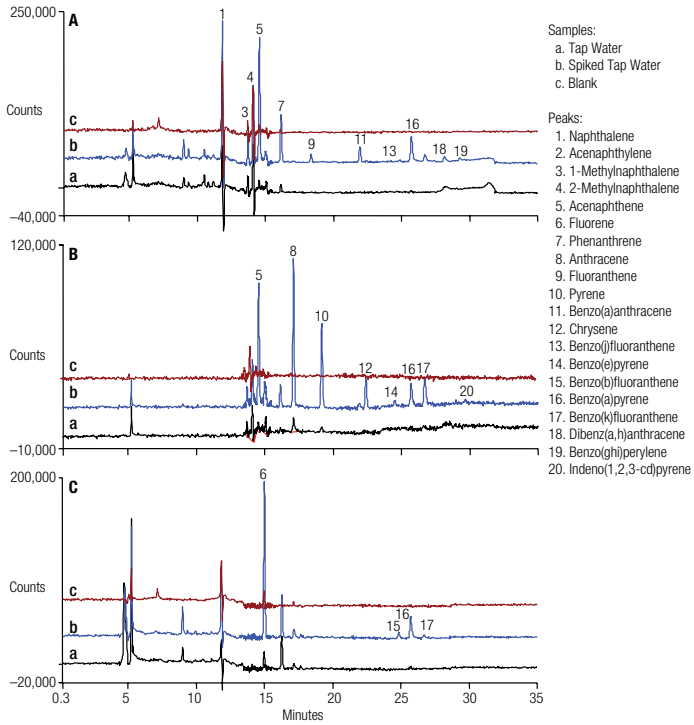

Figure 6. A tap water sample detected by fluorescence using programmed wavelength switching in three parallel channels: (A) Emission_1, (B) Emission_2, (C) Emission_3.

was calculated using the single-sided Student's *t* test method (at the 99% confidence limit). Eight consecutive injections of three reagent water (DI water) samples mixed with 0.1, 1.0, and 10 µg/L of the PAH standard mixtures were used to determine the standard deviation values for calculating MDLs. The calculated MDLs are listed in Table 4.

Analysis of Tap Water Samples

No target analytes were found in the tap water samples. Figure 6 shows chromatograms of a tap water sample detected with fluorescence using three parallel channels. Method recovery was investigated by determining the recoveries in a tap water sample spiked at two concentrations (0.05 and 5 µg/L of each PAH). The recovery range was from 80 to 120%, demonstrating that this on-line SPE HPLC method combined with UV and fluorescence detections provides good selectivity and suitability for the determination of PAHs in water samples.

Conclusion

This work describes an on-line SPE HPLC method with UV absorbance and fluorescence detections for rapid and sensitive determination of 20 PAHs in tap water. The determination is performed on an UltiMate 3000

Dual Gradient LC system controlled by Chromeleon CDS software and combined with a Hypersil Green PAH analytical column. The reduced MDLs for UV and fluorescence detection enabled by on-line SPE using the Acclaim PA2 column provide a convenient method for determining these compounds in drinking and environmental waters using HPLC.

References

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